EIV32025E

ExCell Bio

ResiQuant® Ready-to-use Quantitative *E. coli* DNA Kit (Taqman)

For Research and Manufacturing Use

Not Intended for Diagnostic and Therapeutic Use

User Manual

Catalog Number	CRH00-1101S
	CRH00-1101
	CRH00-1102

PRODUCT DESCRIPTION

The kit is used for the rapid detection of *Escherichia coli* (*E. coli*) host cell DNA in the intermediate, semi-finished and final products of various biologicals and drugs. Uracil N-glycosylase (UNG) is included in the kit to effectively degrade contamination possibly derived from the previous PCR product, thereby greatly reducing the false positive rate. The reagent components include an internal control (IC) and reference dye (ROX). The signal performance of IC allows monitoring of the reaction process to exclude sample interference. ROX is suitable for ABI fluorescence quantitative PCR instrument or other similar equipment and plays the role of optical path correction.

This product is suitable for residual host DNA detection of *E. coli* strains (*E. coli*-K12, *E. coli*-DH5α, *E. coli*-MC1061, *E. coli*-HB101, *E. coli*-XL1-Blue *et al*.). Five well established gradient DNA standards (range from 300 pg/μL to 30 fg/μL) are supplied with the kit as a set of ready-to-use references, simplifying the daily operation and lessening between-run variations.

It is recommended to use the Universal DNA Residual Sample Pretreatment Kit for isolation and purification of the residual DNA from variant samples as in most cases the sample matrix can not be directly used in the detection assay. And for new type of sample matrix, proper suitability study is suggested to be done in advance to make sure the reliability of the test results.

PERFORMANCE, APPLICATION AND RESTRICTION

The kit is suitable for diverse samples from intermediates to final drug products with quantitative detection range from 300 pg/μL to 30 fg/μL.

SPECIFICATION, STORAGE AND TRANSPORTATION REQUIREMENT

Components	CRH00-1101（50T）	CRH00-1102（100T）	CRH00-1101S（50T）
E. coli-Std1	150 µL	250 µL	150 µL
E. coli -Std2	150 µL	250 µL	150 µL
E. coli -Std3	150 µL	250 µL	150 µL
E. coli -Std4	150 µL	250 µL	150 µL
E. coli -Std5	150 µL	250 µL	150 µL
DNA Dilution Buffer	4 mL	4 mL × 2	4 mL
2× *E. coli* qPCR Mix	750 µL	750 µL × 2	750 µL
6× *E. coli* Detection Mix	250 µL	500 µL	250 µL

Storage condition: -40℃ to -18℃.

Validity: 12 months under specified storage conditions.

Transportation: Dry ice.

Applicable instrument: ABI 7500, Bio Rad CFX-96, Agilent MX 3000P *et al*.

EXPERIMENTAL PREPARATION

Instruments and reagents needed

1. Real-time qPCR instrument (FAM and HEX/VIC channels must be included. If "reference" option is available, please select "ROX").
2. Special pipettes and compatible low-retention pipette tips with filter elements.
3. Low-retention microtubes and PCR strips (qPCR-compatible).
4. Lab coats, disposable gloves, face masks, and other PPE.

Division of experimental area

The following division of experimental area is recommended to avoid cross-contamination:

1. Reagent Preparation Area: A physically isolated space (e.g., laminar flow hood) dedicated to the reagents preparation except for samples.
2. Sample Preparation Area: A segregated workspace for nucleic acid extraction, dilution, and other

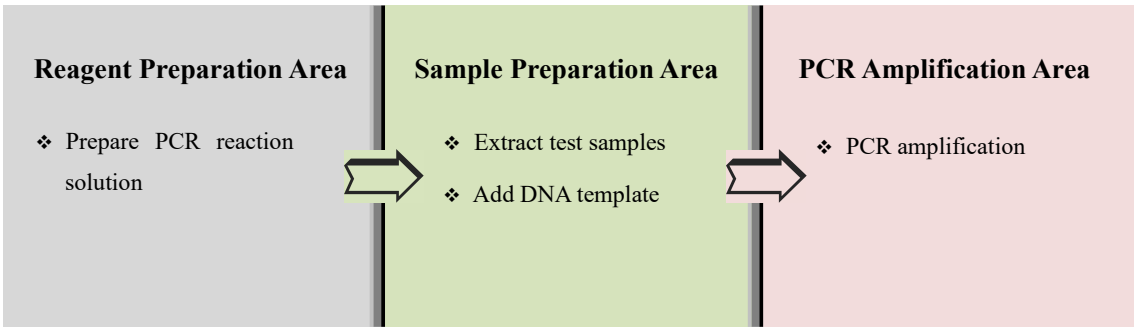
sample-handling steps.

3. PCR Amplification Area: A region for PCR amplification that is independent to the first two regions.

EXPERIMENTAL PROCEDURE

Abbr.	Name	Description
NTC	No Template Control	Negative control
NEG	Negative Extraction Control	Pretreated, negative samples
TS	Test Sample	Sample to be tested
ERC	Extraction Recovery Control	Spiked samples

Operation process



Preparation of PCR reaction solution (Reagent Preparation Area)

For the first use, thaw all components and briefly centrifuge at 1000 × g for 10 sec to ensure reagents are collected at the tube bottom.

1. Determine the number of test samples and controls.
2. Reaction number = (five standard dilutions + one NTC + one NEG + TS + ERC) × 3.
3. Move 6× *E. coli* Detection Mix, 2× *E. coli* qPCR Mix to room temperature and briefly centrifuge.
4. Prepare a PCR reaction mix using the reagents and volumes shown in the table below. Add 20 µL PCR reaction mix to each well (Maintain at 2℃ to 8℃ prior to reaction initiation).

Reagents	Volume for single reaction
2× *E. coli* qPCR Mix	15 µL
6× *E. coli* Detection Mix	5 µL
Total	**20 µL**

Note: *Use 10% excess volume to compensate for pipetting losses.*

Sample preparation (Sample Preparation Area)

DNA extraction:

It is recommended to use Universal DNA Residual Sample Pretreatment Kit to extract host-cell DNA from the samples.

Prepare the standard curve:

1. Move the tubes of *E. coli*-Std1 ~ *E. coli*-Std5 from the freezer and thaw at room temperature. Vortex gently and then briefly centrifuge.

Prepare the PCR plate (Sample Preparation Area)

Plate layout:

	1	2	3	4	5	6	7	8	9	10	11	12
A	NTC	NTC	NTC		TS1	TS1	TS1		TS1 ERC	TS1 ERC	TS1 ERC	
B					TS2	TS2	TS2		TS2 ERC	TS2 ERC	TS2 ERC	
C					TS3	TS3	TS3		TS3 ERC	TS3 ERC	TS3 ERC	
D	ST5	ST5	ST5									
E	ST4	ST4	ST4									
F	ST3	ST3	ST3						NEG	NEG	NEG	
G	ST2	ST2	ST2									
H	ST1	ST1	ST1									

1. Add 10 μL each of DNA template to the corresponding wells.
2. Carefully cover the PCR stripe and briefly centrifuge.

PCR amplification (PCR Amplification Area)

The following steps take the ABI 7500 fluorescence quantitative PCR instrument as an example:

1. Log in, and click “New Experiment” in the upper-left corner of the screen.
2. Enter the name of the experiment, Select “7500 (96 wells), “Quantitation-Standard Curve”, “TaqMan® Reagents” and “Standard”.
3. Click “Plate Setup”, and choose “FAM” as reporter and “None” as quencher. Another reporter target is "VIC", with "None" as the quencher. Add or change sample names if necessary.
4. Click “Assign Targets and Samples”, and set samples, NTCs, and standards in corresponding positions of the plate.

   Select “ROX” in the “Select the dye to use as the passive reference” column.

   Set up the standard serial dilutions with the following steps: (1) Click “Define and Setup Standards” (2) Enter “300” in blank after “Starting Quantity” (3) Choose “1:10” in “Serial Factor” (4) Select and arrange

wells for the standards (5) Click “apply”.

5. Click “Run Method”, and set “Reaction Volume Per Well” as 30 μL, then set up the reaction procedure according to the following table:

Step		Temperature（℃）	Time（s）	Cycles
1	UNG Incubation	37	300	1
2	Initial Denaturation	95	300	1
3	Denaturation	95	15	40
	Annealing & Extension (Fluorescence Collection)	60	60	
The channel for *E. coli*: FAM； The channel for IC: HEX/VIC				

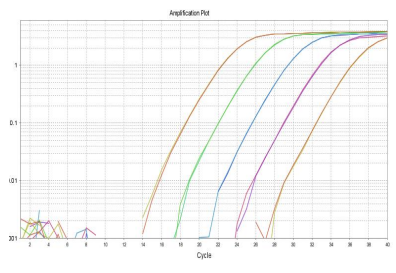
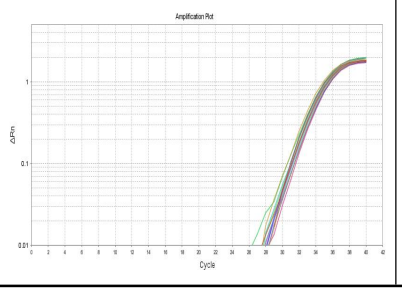
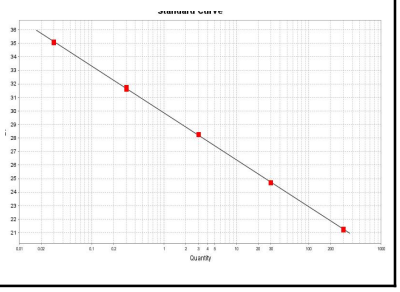
6. After all settings are complete, click the green “Start Run” button.
7. After the run completes, select “Analysis” in the left.
8. Set the threshold to “Auto”, verify that the amplification curves exhibit normal sigmoidal shapes.
9. The slope, intercept and R^2 of the standard curve are shown on the Standard Curve interface.

Quality control

1. $R^2 \geq 0.98$, the slope of the standard curve should be between -3.60 to -3.10, and the amplification efficiency should be between 90% to 110%.
2. The Ct of IC：CV ≤ 5%.
3. The analysis parameters should be set according to the software used, and this process can be automated by the instrument. NTC, NEG Ct ≥ 38 or No Ct.

Description of inspection results

Reference example

FAM	HEX/VIC	Standard Curve
		

Result judgment

The△Ct (HEX/VIC) in the table is the difference between the Ct of sample and the mean Ct of the calibrators,

C_{Sample} represents the concentration of the test sample:

FAM	HEX/VIC	Description	Report
$Ct_{Sample} < Ct_{Std1}$	/	C_{Sample}＞300 pg/μL, samples with concentrations above the ULOQ must be diluted to fall within the quantifiable range and reanalyzed	/
$Ct_{Std1} \leq Ct_{Sample} \leq Ct_{Std5}$	△Ct ＜ -1	The reaction fluid is not uniformly mixed or there is matrix effect	/
	-1≤△Ct≤1	For samples within the quantifiable range, calculate concentrations via the standard curve	Calculate the concentration
	△Ct ＞ 1	The reaction fluid is not uniformly mixed or there is matrix effect	/
$Ct_{Sample} > Ct_{Std5}$ or No Ct	/	Sample concentration below LLOQ	C_{Sample} ＜ 30 fg/μL

Cautions

1. Wear disposable gloves, masks, and a clean lab coat.
2. Use calibrated pipettes (certified within the last 12 months).
3. Use low-retention filter pipette tips.
4. Use dedicated pipettes, pipette tips, and related equipment for each experimental area.
5. Vortex and briefly centrifuge (1000 × g, 10 sec) PCR solutions to collect reagents at the tube bottom.
6. Open and close all reagent/reaction tubes carefully to prevent cross-contamination.
7. Load samples in the following order: NTC, NEG, TS, and ERC.
8. Use separate pipettes for NTC, samples, and DNA template transfer to prevent contamination.
9. Avoid carrying PCR products into reagent preparation or sample preparation areas.
10. All workbenches and instruments must be cleaned with 75% alcohol after use.

DISCLAIMER

1. The product should be used according to the instructions in the manual. If the experimenter fails to operate according to the instructions, our company will not be responsible for any deviation in product performance caused by this.
2. The product is only used for scientific research and commercial production, and is not suitable for clinical diagnosis and treatment. Otherwise, all consequences arising shall be borne by the experimenter, and our company shall not be responsible.