

# **ExCell Bio**

# OptiVitro® CHO Serum-free Basal Medium TransExp CE06

For Research and Manufacturing Use

Not Intended for Diagnostic and Therapeutic Use

# User Manual Catalog Number CE000-N052 Catalog Number CE000-N053 Catalog Number CE000-N054 Catalog Number CE000-N055



## | PRODUCT DESCRIPTION

OptiVitro® CHO Serum-free Basal Medium TransExp CE06 is a state-of-the-art, animal component-free and chemically-defined medium that is specifically designed for the high-density culture transfection of suspended CHO-K1 and CHO-S etc., especially suitable for ExpiCHO-S cells. This product does not require additional glutamine. It can be used together with OptiVitro® CHO Serum-free Feed Medium TransExp CA06 (Catalog no.CA000-N03#) to promote protein production.

# | SPECIFICATION, STORAGE AND TRANSPORTATION

### REQUIREMENT

Product Name	Cat.#	Specification	Storage	Transportation	Shelf Life
OptiVitro® CHO Serum-free Basal Medium TransExp CE06	CE000-N052	1000m L Liquid	2-8 °C Protect From Light	<25 °C Protect From Light	12 months
	CE000-N053	1 L Powder		410.9C	24 months
	CE000-N054	10 L Powder	10 L 2-8 °C Protect  Powder Dark and Dry		
	CE000-N055	100 L Powder		From Light	

# PERFORMANCE, APPLICATION AND RESTRICTION

- Please make sure to store the cell culture medium in a light-protected environment, avoid fluorescent lamps or other lamplight exposure, and better to use colored packaging bags in the refrigerator or warehouse.
- 2. During the transportation of products, try to avoid the impact of fluorescent lamps or other lamplight exposure on the appearance of the product, resulting in appearance discoloration.
- 3. During the transportation of the product to the clean area, it is essential to carry out a cleaning and sterilization process. The sterilization method may involve disinfectant wiping, and not utilize UV irradiation for sterilization.

[ Note ]: When passing through transfer windows equipped with UV sterilization, remember to proactively

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switch off the UV lamp inside the transfer window.

INSTRUCTION FOR USE

**Medium preparation** 

1. Measure 80% of the final volume WFI or distilled water in a clean vessel.

2. Add 23.52g/L OptiVitro® CHO Serum-free Basal Medium TransExp CE06 powder slowly to the water,

mix for 20 minutes.

3. Adjust the pH to 8.0-8.1 with 5 N NaOH (about 3.6 mL). After adjusting, continue stirring for an

additional 30 minutes.

4. Adjust the pH to 7.0-7.1 with 6 N HCl (about 1.6 mL). After adjusting, continue stirring for an

additional 10 minutes.

5. Add 2.2g/L NaHCO<sub>3</sub> and mix for 10-15 minutes, adjust the pH to 6.9-7.1.

6. QS to final production volume and mix for 5 minutes.

7. Measure the final pH (6.90-7.50).

 $8\,.\,$  Measure the osmolality (275-305 mOsm/kg).

9. Sterilize immediately by membrane filtration (< 0.22 microns), and store at 2 to 8°C.

**Cell Culture** 

Suggested culture condition, Temperature: 37°C, RH:80%, CO<sub>2</sub>:5%, 120rpm. According to the cell growth,

it can be passed every 2-4 days when the living cell density reaches 4.0-6.0×10<sup>6</sup> cells/mL, and the passage

density is 0.2-0.3×10<sup>6</sup> cells/mL.

**Pre-transfection Preparation** 

1. Cell Maintenance

(1) Perform at least three consistent passages following cell recovery.

(2) Ensure cell viability exceeds 90% before proceeding with transfection.

2. Cell Seeding (Day -1)

(1) Resuspend cells in fresh OptiVitro® CHO Serum-free Basal Medium TransExp CE06.

(2) Initial seeding density:  $3.5 \times 10^6$  cells/mL.

[Note]: Omitting centrifugation may result in reduced protein expression.

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### Transfection Procedure (Day 0)

### **Cell Preparation**

- 1. Adjust cell suspension to 18 mL using fresh OptiVitro® CHO Serum-free Basal Medium TransExp CE06.
  - 2. Target parameters:
  - (1) Total cell count:  $1.2 \times 10^8$  cells.
  - (2) Final transfection density:  $6 \times 10^6$  cells/mL in 20 mL culture.

### **PEI/DNA Complex Formation**

- 1. Transfection Parameters:
- (1) Culture volume: 20 mL.
- (2) Cell density:  $6 \times 10^6$  cells/mL.
- (3) DNA concentration: 1.5 μg/mL.
- (4) DNA:PEI ratio: 1:4.
- 2. Complex Preparation Steps:
- a) PEI Max Solution:
- (1) Dilute 120 μg PEI Max in OptiVitro® CHO Serum-free Basal Medium TransExp CE06 to 1 mL.
- (2) Incubate at room temperature for 5 minutes.
- b) DNA Solution:
- (1) Dilute 30 µg DNA in OptiVitro® CHO Serum-free Basal Medium TransExp CE06 to 1 mL.
- c) Complex Formation:
- (1) Add PEI Max solution to DNA solution.
- (2) Mix thoroughly.
- (3) Incubate at room temperature for 10 minutes.
- 3. Transfection
- (1) Add 2 mL PEI/DNA complex to cell suspension.
- (2) Maintain continuous flask agitation during addition for optimal mixing.

### **Post-transfection Process**

- 1. Initial Feed (18-24 hours post-transfection):
- (1) Add 5% volume OptiVitro® CHO Serum-free Feed Medium TransExp CA06.

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(2) Supplement with glucose to final concentration of 22 g/L.

(3) Shift culture temperature to 33°C.

2. Feed Strategy Options:

a) Basic Strategy (for standard protein yields):

(1) Single feed at 18-24 hours post-transfection (as described above).

(2) Recommended when simplified operation is preferred.

b) Enhanced Strategies (for maximizing protein yields):

(1) Option 1: Two-step feeding.

Add 5% OptiVitro® CHO Serum-free Feed Medium TransExp CA06 on D1 and D3.

(2) Option 2: Three-step feeding.

Add 5% OptiVitro® CHO Serum-free Feed Medium TransExp CA06 on D1, D3, and D6.

[ Note ] : Additional feeding strategies should be implemented based on protein yield requirements and operational considerations.

3. Optional Enhancement

(1) Add OptiVitro® protein expression enhancer (Catalog no. M10141#) on D1.

(2) Refer to Table 1 for specific addition volumes.

Harvest

1. Standard harvest time: Day 10 post-transfection.

2. Earlier harvest may be considered based on:

(1) Target protein characteristics.

(2) Cell viability status.

[Note]

The aforementioned transfection techniques are intended for informational purposes solely. To achieve the most favorable transfection parameters for various CHO cells, a Design of Experiments (DoE) approach can be utilized for establishing the optimal experimental design, encompassing cell density, DNA quantity, and DNA to PEI ratio.

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Table 1. Recommended dosage for various transfection specifications

Cell culture vessel	125 mL	500 mL	1 L		
Amount of cell (x 10 <sup>6</sup> cells/mL)	120	600	1200	cell density  6x 10 <sup>6</sup> cells/mL	
OptiVitro® CHO Serum-free Basal Medium TransExp CE06 (mL)	18	90	180	Initial culture volume	
DNA diluent (mL)	1	5	10	Initial culture volume	
PEI diluent (mL)	1	5	10		
DNA (μg)	30	150	300	DNA.DEL_1.4	
PEI Max (μg)	120		1200	DNA:PEI=1:4	
OptiVitro® CHO Serum-free Feed	1	5	10	5% of the initial	
Medium TransExp CA06 (mL)	Medium TransExp CA06 (mL)		10	transfection volume	
OptiVitro® Protein expression 0.02		0.1	0.2	0.1% of the initial	
enhancer(mL)	0.02	0.1	0.2	transfection volume	
Final culture system (mL)	~21	~105	~210		

Table 2. Related product number

Product Name	Cat.#	Specification	
	CA000-N032	100 mL Liquid	
OptiVitro® CHO Serum-free Feed Medium	CA000-N033	1000mL Liquid	
TransExp CA06	CA000-N034	1 L Powder	
	CA000-N035	10 L Powder	
OptiVitro® Glucose solution	M101382C	100 mL Liquid	
OutiVitus® Protein averaging anhances	M101412C	1 mL Liquid	
OptiVitro® Protein expression enhancer	M101413C	5 mL Liquid	

# **DISCLAIMER**

- 1. The product should be used according to the instructions in the manual. If the experimenter fails to operate according to the instructions, our company will not be responsible for any deviation in product performance caused by this.
- 2. The product is only used for scientific research and commercial production, and is not suitable for clinical diagnosis and treatment. Otherwise, all consequences arising shall be borne by the experimenter, and our company shall not be responsible.

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